

Four New Records of Monogonont Rotifers (Rotifera: Monogononta) from Korea

Hee-Min Yang, Gi-Sik Min*

Department of Biological Sciences and Bioengineering, Inha University, Incheon 22212, Korea

ABSTRACT

Four monogonont rotifers, *Filinia hofmanni* Koste, 1980, *Lecane pusilla* Haring, 1914, *Mikrocodides chlaena* (Gosse, 1886), and *Proales fallaciosa* Wulfert, 1937, were newly recorded in Korea. The genera *Mikrocodides* Bergendal, 1892 and *Proales* Gosse, 1886 were recorded for the first time in Korea. *Mikrocodides chlaena* and *Proales fallaciosa* were found from soil samples and are both soft-bodied species. *Filinia hofmanni* has previously been recorded mainly in Europe, and this is its first record in Asia. *Lecane pusilla* is the 24th lecanid rotifer recorded in Korea, and its morphological characteristics are consistent with previous research of *L. pusilla*. We have provided the morphological diagnoses of the four Korean specimens in this study, along with the partial sequences of mitochondrial cytochrome *c* oxidase subunit I (COI) gene of three species.

Keywords: *Filinia*, *Lecane*, *Mikrocodides*, Monogononta, *Proales*

INTRODUCTION

The monogonont rotifers in Korea have continuously been recorded by several researchers since the report by Hada in 1936 (Yamamoto, 1953; Turner, 1986; Song and Kim, 1989; Chung et al., 1991; Kim et al., 1993). To date, 323 rotifer species have been recorded in Korea, of which 200 are monogonont rotifers (National Institute of Biological Resources, 2021).

Among the monogonont rotifers recorded in Korea, there was a research bias for some specific taxa, especially for those with hard lorica or those which do not serious contraction or distorted due to fixative solutions, such as *Brachionus* Pallas, 1766, *Keratella* Bory de St. Vincent, 1822, *Lecane* Nitzsch, 1827, *Lepadella* Bory de St. Vincent, 1826, and *Trichocerca* Lamarck, 1801. In addition, some genera, especially those with soft bodies, were rarely studied in Korea. For example, the family Dicranophoridae Haring, 1913 and Notommatidae Hudson & Gosse, 1886 contain 236 and 268 species, respectively (Jersabek and Leitner, 2013), but in Korea only 7 and 14 species, respectively, have been recorded.

Because of this research bias, we tried to find the monogonont rotifers that have not been studied much in Korea, especially the soft-bodied species. In Korea, the discovery of

various taxa of monogonont rotifers is expected. The unique geographic and climatic characteristics of more than 60% of land is made up of mountains, land is surrounded by sea containing thousands of islands, and four distinct seasons with diverse climate patterns (Republic of Korea, 2014), create various habitats in Korea. Therefore, it is necessary to investigate various habitats by season to reveal the diversity of rotifers in Korea. In addition, since rotifers inhabit various habitat, it is necessary to apply various collection methods and tools.

In this study, we identified four monogonont rotifers that were newly recorded in Korea: *Filinia hofmanni* Koste, 1980; *Lecane pusilla* Haring, 1914; *Mikrocodides chlaena* (Gosse, 1886); and *Proales fallaciosa* Wulfert, 1937. This is the first record of the genera *Mikrocodides* Bergendal, 1892 and *Proales* Gosse, 1886 in Korea. *Mikrocodides chlaena* and *P. fallaciosa* are both soft-bodied species and considered to have a cosmopolitan distribution. In case of *F. hofmanni* and *L. pusilla*, this is the seventh and the 24th record of the respective genera from Korea. *Filinia hofmanni* has been considered a European species, and this is the first record of it in Asia. Here, we provide the diagnoses of the four Korean specimens and the partial mitochondrial cytochrome *c* oxidase subunit I (COI) sequences of three species.

© This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

***To whom correspondence should be addressed**

Tel: 82-32-860-7692, Fax: 82-32-874-6737
E-mail: mingisik@inha.ac.kr

MATERIALS AND METHODS

Of the four species, *F. hofmanni* and *L. pusilla* were collected from ponds using a 50 µm plankton net, while *M. chlaena*, and *P. fallaciosa* were collected and isolated from soil samples. The soil samples were air-dried in laboratory for a few weeks and rewetted using mineral water in a plant culture dish (SPL Life Science, Korea). The hatching rotifers were isolated into new plant culture dishes and cultured at room temperature.

For the morphological identification, we observed the rotifers under optical microscope (DM2500; Leica, Germany), and identified them based on the Koste and Voigt (1978), Koste and Shiel (1990a, 1990b), Segers (1995), Sanoamuang (2002), and Jersabek and Leitner (2013). Trophi preparation for scanning electric microscopy (SEM) was done following the method described by De Smet (1998). SU8010 and S-4200 (Hitachi, Japan) were used for SEM at 10–15 accelerating voltage. All examined specimens have been deposited in the National Institute of Biological Resources (NIBR).

DNA of three rotifer species were extracted using a LaboPass Tissue Genomic DNA Isolation Kit Mini (Cosmo Genetech, Korea) following the manufacturer's protocol. In case of *L. pusilla*, DNA could not be extracted because it was stored in a formalin-preserved sample. Polymerase chain reaction (PCR) was performed using two COI primers, LCO1490/HCO2198 (Folmer et al., 1994) and 30F/885R (Zhang et al., 2021). The PCR conditions for the Folmer primer were as follows: 95°C for 2 min for the initial denaturation; followed by 40 cycles at 95°C for 15 s, 42°C for 30 s, and 72°C for 1 min; and a final extension at 72°C for 5 min. The PCR conditions for 30F/885R were the same as that for the Folmer primer, except the annealing temperature of 51°C. PCR products were visualized by 1% agarose gel electrophoresis, and purified using LaboPass PCR Purification Kit (Cosmo Genetech). DNA sequencing was performed using PCR primers and ABI 3700 DNA Analyzer (Applied Biosystems, USA) at Macrogen, Korea.

The sequences were assembled and trimmed using Geneious 8.1.9 (<https://www.geneious.com>), and the intra-specific genetic variation were calculated using MEGA11 (Tamura et al., 2021).

SYSTEMATIC ACCOUNTS

¹*Phylum Rotifera Cuvier, 1817

²*Class Eurotatoria De Ridder, 1957

³*Subclass Monogononta Plate, 1889

⁴*Order Flosculariaceae Harring, 1913

Family Trochosphaeridae Harring, 1913

⁵*Genus *Filinia* Bory de St. Vincent, 1824

⁶**Filinia hofmanni* Koste, 1980 (Figs. 1B, 2A, B)

Filinia cf. *longiseta* Hofmann, 1979.

Filinia hofmanni: Koste, 1980: 240–245; Jersabek, 1996: 83; Sanoamuang, 2002: 244–245.

Material examined. Korea: 5 individuals, Incheon, Pond at Inha University, 37°27'00"N, 126°39'22"E, 19 Dec 2019.

Diagnosis. Body 130–160 µm in length, sac-shaped or oviform. Two lateral setae and one caudal seta present, lateral setae equal in length and inserted near corona. Lateral setae 260–300 µm, 2–3 times longer than body length. One movable caudal seta inserted ventrally. Caudal seta 137–170 µm in length. Distance from posterior end to caudal seta insertion is 15–25 µm. Two red eyes on head. Vitellarium with 14 nuclei. Trophi malleoramate type. Fulcrum straight, thin in dorso ventral view, broad and axe-shaped in lateral view. Rami inner margin with scleropili. Basal chamber of rami within thin wall structure, divide into the basal ramus chamber and the subbasal ramus chamber. Manubria crescent-shape, divided to three chambers. Unci located on the inner margin of manubria, containing 12/13 unci teeth.

Distribution. Austria, Belgium, Germany, Spain, Korea.

Remarks. *Filinia hofmanni* is similar to *F. longiseta* in morphological characteristics: sac-shaped body, movable caudal seta and the position of caudal seta insertion (Sanoamuang, 2002). The biggest difference between the two species is the number of teeth on unci. The number of unci teeth is a species-specific characteristic in the genus *Filinia* because it remains constant regardless of the environmental factors or the developmental stages (Sanoamuang, 1993). The European *F. hofmanni* have much fewer unci teeth than *F. longiseta*: 13–14/14–16 vs. 18–20/20–22 (Sanoamuang, 2002). The Korean specimen also has fewer teeth than *F. longiseta*: 12/13, which has one to three lesser number of teeth on each side than European *F. hofmanni*. The temperature of habitats was also different for *F. hofmanni* and *F. longiseta*. *Filinia longiseta* was recorded as warm-stenotherm (23–31°C), whereas *F. hofmanni* was recorded as cold-stenotherm (4–6°C) (Sanoamuang, 2002). The Korean specimen was also collected from cold water only (4–5°C).

Deposition. NIBRIV0000879590.

Molecular data. The partial COI sequences were obtained from four specimens. Intra-specific genetic distance was 0.0–1.4% (630 bp) (GenBank accession numbers: OM142482–OM142485).

Korean name: ¹*윤형동물문, ²*진원충강, ³*단소아강, ⁴*소화윤충목, ⁵*삼지윤충속, ⁶*호프만삼지윤충 (신칭)

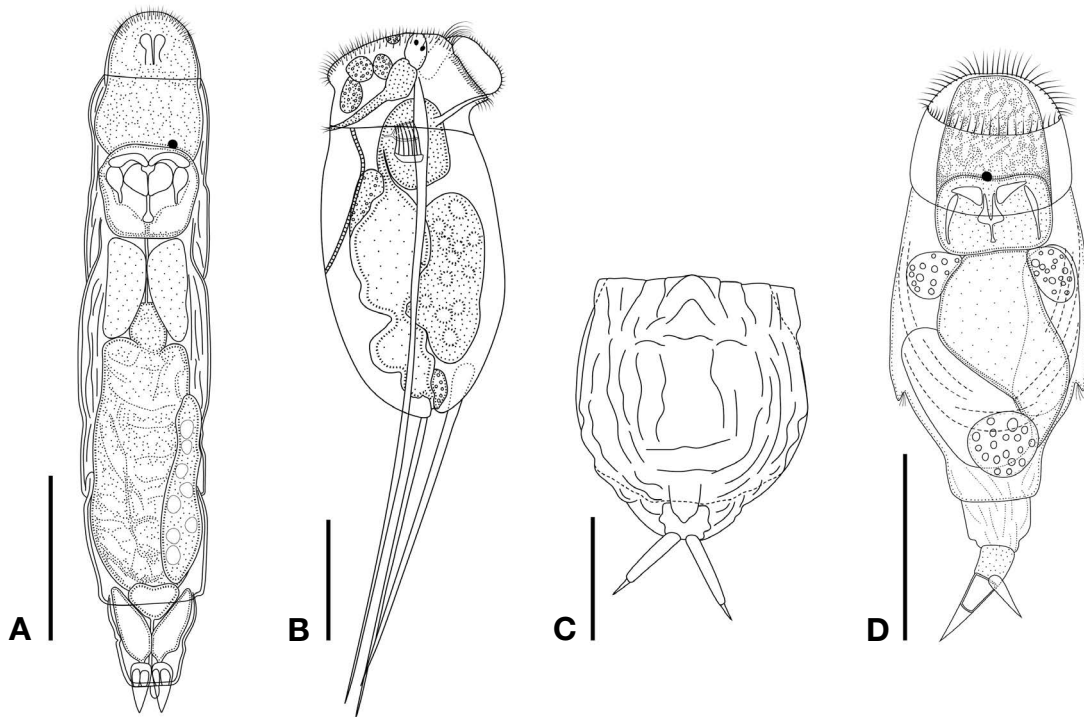


Fig. 1. Line drawing of rotifers. A, Dorsal view of *Proales fallaciosa*; B, Lateral view of *Filinia hofmanni*; C, Ventral view of *Lecane pusilla*; D, Dorsal view of *Mikrocodides chlaena*. Scale bars: A, B, D=50 μ m, C=25 μ m.

¹*Order Ploima Hudson & Gosse, 1886

²*Family Lecanidae Remane, 1933

³*Genus *Lecane* Nitzsch, 1827

⁴**Lecane pusilla* Haring, 1914 (Fig. 1C)

Lecane pusilla: Haring, 1914: 541; Haring and Myers, 1926: 369; Kutikova, 1970: 449–450; Chengalath et al., 1973: 20; Koste and Voigt, 1978: 216; Koste and Shiel, 1990a: 31; Segers, 1995: 84.

Lecane nana Donner, 1954: 85–86.

Material examined. Korea: 3 individuals, Jeju-do, Pond in Seogwipo-si, 33°25'13"N, 126°50'42"E, 25 Jun 2021.

Diagnosis. Antero-lateral corner of lorica angulated. Dorsal plate anteriorly narrower than ventral plate. Anterior margin of dorsal plate flat. Middle of ventral plate narrower than dorsal plate. Length of dorsal plate is 50 μ m, and that of ventral plate is 55 μ m. Width of dorsal plate is 45 μ m, and that of ventral plate is 42 μ m. Both dorsal and ventral lorica ornamented, showing a pleated pattern. Posterior margin of foot plate round. Foot pseudosegment squircle shaped with a lateral extension in middle, not projecting from ventral plate. Toes short, 20 μ m in length, and completely separated.

Tips of toes bearing short claws, 4–5 μ m in length.

Distribution. Cosmopolitan.

Remarks. *Lecane pusilla* has been recorded in all continents except the polar regions (Haring, 1914; Haring and Myers, 1926; Donner, 1954; Kutikova, 1970; Chengalath et al., 1973; De Smet, 1989; Koste and Shiel, 1990a; Sanoamuang et al., 1995). The morphological characteristics of *L. pusilla* can be confused with the following lecanid species: *L. aeganea* Haring, 1914, *L. doryssa* Haring, 1914, *L. inopinata* Haring & Myers, 1926, *L. sagula* Haring & Myers, 1926, *L. subtilis* Haring & Myers, 1926, and *L. undulata* Hauer, 1938, but can be distinguished by its small body size, completely separated toes, short claws, shape of foot pseudosegment, and the foot pseudosegment not projecting from ventral plate (Segers, 1995). The morphological characteristics of Korean *L. pusilla* are consistent with previous research.

Deposition. NIBRIV0000895314.

⁵*Family Epiphanidae Haring, 1913

⁶*Genus *Mikrocodides* Bergendal, 1892

⁷**Mikrocodides chlaena* (Gosse, 1886) (Figs. 1D, 2E, F)
Stephanops chlaena Hudson and Gosse, 1886: 76.

Korean name: ¹*유영목, ²*술잔윤충과, ³*술잔윤충속, ⁴*작은술잔윤충 (신칭), ⁵*물윤충과, ⁶*등주름윤충속 (신칭), ⁷*외발톱등주름윤충 (신칭)

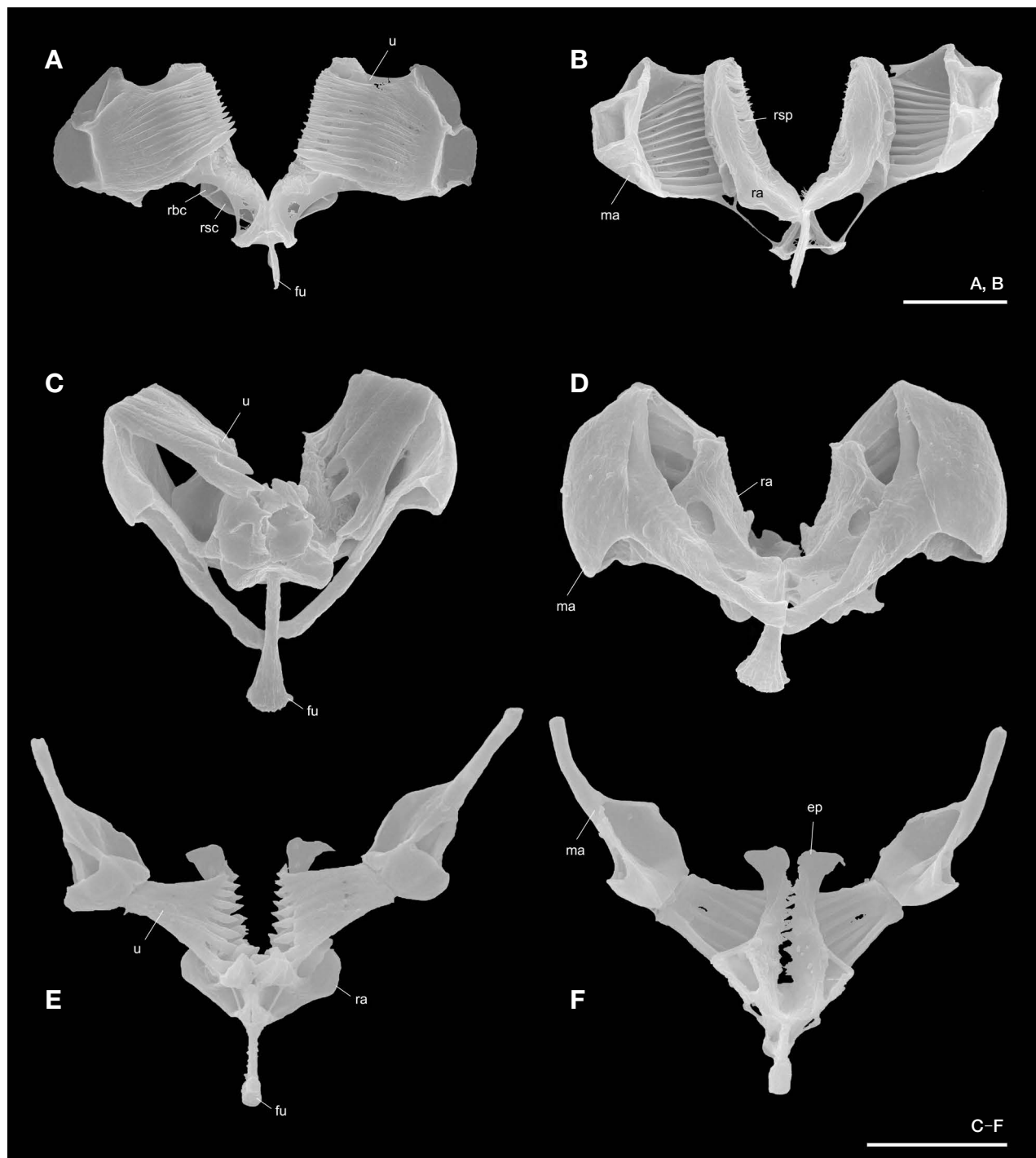


Fig. 2. Trophi observed by scanning electric microscopy. A, B, *Filinia hofmanni*; A, Ventral view; B, Dorsal view; C, D, *Proales fallaciosa*; C, Ventral view; D, Dorsal view; E, F, *Mikrocodides chlaena*; E, Ventral view; F, Dorsal view. ep, epipharynx; fu, fulcrum; ma, manubrium; ra, ramus; rbc, ramus basal chamber; rsc, ramus subbasal chamber; rsp, ramus scleropili; u, uncus. Scale bars: A-F= 10 μ m.

Rhinops orbiculodiscus Thrope, 1891: 304.
Mikrocodides dubius Bergendal, 1892: 34.
Mikrocodides orbiculodiscus Jennings, 1894: 8.

Mikrocodides chlaena: Weber, 1898: 369; Wang, 1961: 108;
 Kutikova, 1970: 500; Koste and Voigt, 1978: 61; Koste
 and Shiel, 1987: 966; Jersabek, 2003: 62.

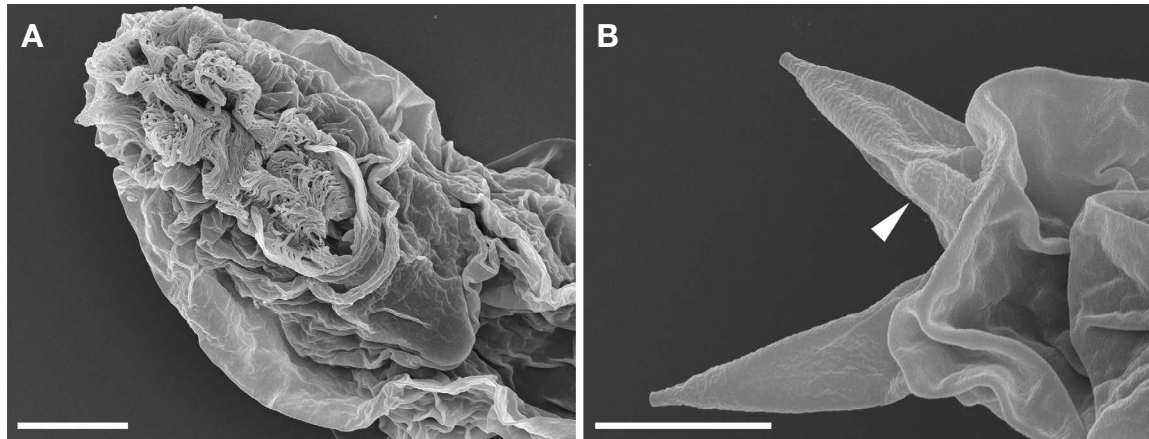


Fig. 3. *Proales fallaciosa* observed by scanning electric microscopy. A, Ventral view of corona; B, Toes and dorsal papilla (arrowhead). Scale bars: A = 10 μ m, B = 5 μ m.

Material examined. Korea: 5 individuals, Gyeongsangnam-do, Soil sample from Gimhae-si, 35°19'01"N, 128°48'10"E, 13 May 2020.

Diagnosis. Body flexible and illoricated, 150–160 μ m in length. Corona short, located on the anterior margin of head. Head large, conspicuously divided from trunk. Trunk cylindrical, with lateral projections from two thirds of the trunk. Dorsal part of lorica with distinct longitudinal folds. Two toes completely separated. Toes of different lengths, 20 and 16 μ m. Long toe pointed backwards and short toe pointed upwards. One red eyespot on brain. Trophi malleate, 17 μ m in length. Fulcrum straight, with thick and blunt end. Rami rhomboid with several teeth on the inner margin. Upper part of rami connected to epipharynx. Unci with eight distinct teeth.

Distribution. Cosmopolitan.

Remarks. The genus *Mikrocodides* Bergendal, 1892 comprises of two species, *M. chlaena* and *M. hertha* Wulfert, 1961 (Jersabek and Leitner, 2013). *Mikrocodides chlaena* is easily distinguished from *M. hertha* based on the following characteristics: (1) toes asymmetrical and unique, (2) one cerebral eye, and (3) unci with eight teeth. In several studies on *M. chlaena* (Weber, 1898; Wang, 1961; Kutikova, 1970; Koste and Voigt, 1978; Koste and Shiel, 1987; Jersabek, 2003), the genus name was notated as '*Microcodides*'. We follow the genus name '*Mikrocodides*' based on the original description (Bergendal, 1892) and the standard list of names for use in taxonomy (Segers et al., 2012). Of the five genera of the family Epiphanidae Haring, 1913, two genera *Epiphanes* Ehrenberg, 1832 and *Rhinoglena* Ehrenberg, 1853 have been recorded previously from Korea (National Institute of

Biological Resources, 2021), and this is the first record of the genus *Mikrocodides* in Korea.

Deposition. NIBRIV0000879591.

Molecular data. The partial COI sequences were obtained from three specimens, and the intra-specific distance was zero (663 bp) (GenBank accession numbers: OM142489–OM142491).

¹*Family Proalidae Haring & Myers, 1924

²*Genus *Proales* Gosse, 1886

³**Proales fallaciosa* Wulfert, 1937 (Figs. 1A, 2C, D, 3A, B)

Proales decipiens Weber, 1898: 466.

Proales sordida Haring & Myers, 1922: 605.

Proales tyrphosa Bērziņš, 1948: 315.

Proales fallaciosa: Wulfert, 1937: 65; Wang, 1961: 159; Kutikova, 1970: 496; Koste and Voigt, 1978: 281; Koste and Shiel, 1990b: 135; De Smet, 1996: 70; Fontaneto and Melone, 2003: 258; Jersabek, 2010: 154.

Material examined. Korea: 5 individuals, Gyeongsangnam-do, Soil sample from Sancheong-gun, 35°27'54"N, 127°51'50"E, 19 Jan 2019.

Diagnosis. Body illoricated and flexible, 220–250 μ m in length. Transparent and elongated cylindrically. Corona oblique. Trunk with pseudosegments and several longitudinal folds. Foot short, less than 1/8 of the total body length. Two pseudosegments on the foot. One small dorsal papilla located between two toes. Toes short, 7–8 μ m in length. Tips of toes truncated. One red eyespot located behind the brain, slightly

Korean name: ¹*돌출윤충과, ²*돌출윤충속 (신칭), ³*작은돌기돌출윤충 (신칭)

right from the center in dorsal view. A pair of large and elongated gastric glands. Pedal glands subtriangular shape. Vitellarium with eight nuclei. Trophi virgate. Fulcrum extended straight with fan-shaped distal end. Length of fulcrum 8–9 μm . Inner margin of rami contains one blunt tooth on each side. Manubria broad at the anterior end, narrow and straight at the posterior end. Each uncus with five teeth with gradually decreasing sizes.

Distribution. Cosmopolitan.

Remarks. *Proales fallaciosa* is the first species of the genus *Proales* to be recorded in Korea. Previously, three species of the family Proalidae have been recorded in Korea, namely *Bryceella perpusilla* Wilts, Martinez Arbizu & Ahlrichs, 2010, *B. stylata* (Milne, 1886), and *Proalinopsis caudatus* (Collins, 1872) (see Song, 2015, 2017). The morphological characteristics of the Korean specimen corresponded to the previous studies except the number of unci teeth. De Smet (1996) recorded that the range of the unci teeth number was from 4 to 7, and Kutikova (1970) recorded that the Russian specimen had 6 teeth on the right uncus and 7 teeth on the left. The Australian specimen had been recorded to have 7 teeth on left and 5–6 teeth on right (Koste and Shiel, 1990b). The Korean specimen has 5 teeth on each side. Of the 48 species in the genus *Proales*, *P. fallaciosa* is the most similar to *P. gigantea* (Glascotte, 1893) and *P. ornata* Myers, 1933. However, *P. fallaciosa* is distinguished from the two species based on the following characteristics: (1) dorsal papilla of *P. fallaciosa* is simple and smaller than *P. gigantea*, (2) the length of two pseudosegments of foot is almost similar in *P. fallaciosa*, while that of the last segment of *P. ornata* is twice that of the penultimate segment, and (3) *P. ornata* has lateral protrusions beside the dorsal papilla, while *P. fallaciosa* has no lateral protrusions.

Deposition. NIBRIV0000879589.

Molecular data. The partial COI sequences were obtained from three specimens, and the intra-specific distance was zero (657–660 bp) (GenBank accession number: OM142486–OM142488).

ORCID

Hee-Min Yang: <https://orcid.org/0000-0001-6288-1534>

Gi-Sik Min: <https://orcid.org/0000-0003-2739-3978>

CONFLICTS OF INTEREST

Gi-Sik Min, a contributing editor of the Animal Systematics, Evolution and Diversity, was not involved in the edito-

rial evaluation or decision to publish this article. Remaining author has declared no conflicts of interest.

ACKNOWLEDGMENTS

This work was supported by a grant from the National Institute of Biological Resources (NIBR), funded by the Ministry of Environment (MOE) of the Republic of Korea (NIBR 202002204).

REFERENCES

- Bergendal D, 1892. Beiträge zur Fauna Grönlands. Ergebnisse einer im Jahre 1890 in Grönland vorgenommenen Forschungsreise. I. Zur Rotatorienfauna Grönlands. Berlingska Boktryckeri, Lund, pp. 1-180.
- Bërziņš B, 1948. Einige neue Notommatidae-Arten (Rotatoria) aus Schweden. Hydrobiologia, 1:312-321.
- Chengalath R, Fernando CH, Koste W, 1973. Rotifera from Sri Lanka (Ceylon) 2. Further studies on the Eurotatoria including new records. Bulletin of the Fisheries Research Station of Sri Lanka (Ceylon), 24:29-62.
- Chung EC, Yoo HB, Kim SY, 1991. Freshwater Rotifera of Korea I. Family Lecanidae (Rotifera: Monogononta). Korean Journal of Limnology, 24:207-225.
- De Smet WH, 1989. Contributions to the rotifer fauna of the Bas-Zaïre. 1. The rotifers from some small ponds and a river. Biologisch Jaarboek Dodonaea, 56:115-131.
- De Smet WH, 1996. Guides to the identification of the micro-invertebrates of the continental waters of the world. 9. Rotifera Vol. 4: The Proalidae (Monogononta). SPB Academic Publishing bv, Amsterdam, pp. 1-102.
- De Smet WH, 1998. Preparation of rotifer trophi for light and scanning electron microscopy. Hydrobiologia, 387/388:117-121. https://doi.org/10.1007/978-94-011-4782-8_17
- Donner J, 1954. Zur Rotatorienfauna Südmährens. Österreichische Zoologische Zeitschrift, 5:30-117.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R, 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology, 3:294-299.
- Fontaneto D, Melone G, 2003. On some rotifers new for the Italian fauna. Italian Journal of Zoology, 70:253-259. <https://doi.org/10.1080/11250000309356526>
- Hada Y, 1936. The plankton of lake Seiko, Suigen in autumn. Journal of Chosen Natural History Society, 21:1-11.
- Harring HK, 1914. Report on Rotatoria from Panama with description of new species. Proceedings of the United States National Museum, 47:525-564. <https://doi.org/10.5479/si.00963801.47-2062.525>

- Harring HK, Myers FJ, 1922. The rotifer fauna of Wisconsin. Transactions of the Wisconsin Academy of Sciences, Arts and Letters, 20:553-662.
- Harring HK, Myers FJ, 1926. The rotifer fauna of Wisconsin. III. A revision of the genera *Lecane* and *Monostyla*. Transactions of the Wisconsin Academy of Sciences, Arts and Letters, 22:315-423.
- Hofmann W, 1979. Untersuchungen zur ökologischen Nische zweier nahverwandter planktischer Rotatorien-Arten des Plußsees. Christian-Albrechts-Universität Kiel, Kiel, pp. 1-139.
- Hudson CT, Gosse PH, 1886. The rotifera; Wheel-Animalcules. Vol. II. Longmans, Green, and Co., London, pp. 1-144.
- Jenning HS, 1894. A list of the Rotatoria of the Great Lakes and of some of the Inland Lakes of Michigan. Bulletin of the Michigan Fish Commission, 3:1-34.
- Jersabek CD, 1996. Verbreitung, Ökologie und Taxonomie von Rädertieren (Rotifera) in alpinen Gewässern der Hohen Tauern und der Nördlichen Kalkalpen. Berichte der naturwissenschaftlich-medizinischen Vereinigung Salzburg, 11:73-145.
- Jersabek CD, 2003. Freshwater Rotifera (Monogononta) from Hawai'i: a preliminary checklist. Bishop Museum Occasional Papers, 74:46-72.
- Jersabek CD, 2010. Mongolian rotifers (Rotifera, Monogononta): a checklist with annotations on global distribution and autecology. Proceedings of the Academy of Natural Sciences of Philadelphia, 159:119-168. <https://doi.org/10.1635/053.159.0108>
- Jersabek CD, Leitner MF, 2013. The Rotifer World Catalog. World Wide Web electronic publication [Internet]. The Author, Accessed 10 Jan 2022, <<http://www.rotifera.hausder-natur.at/>>.
- Kim SE, Chung CE, Yoo HB, 1993. Rotifera from Korean inland waters, VI. *Anuraeopsis*, *Mytilina*, *Trichotria*, *Lophocaris*, *Dipleuchlanis* of Brachionidae (Rotifera: Monogononta). Korean Journal of Limnology, 26:293-303.
- Koste W, 1980. Über zwei Plankton-Rädertiertaxa *Filinia australiensis* n. sp. und *Filinia hofmanni* n. sp., mit Bemerkungen zur Taxonomie der longiseta-terminalis-Gruppe. Genus *Filinia* Bory de St. Vincent, 1824, Familie Filiniidae Bartos, 1959 (Überordnung Monogononta). Archiv für Hydrobiologie, 90:230-256.
- Koste W, Shiel RJ, 1987. Rotifera from Australian inland waters. II. Epiphanidae and Brachionidae (Rotifera: Monogononta). Invertebrate Taxonomy, 7:949-1021. <https://doi.org/10.1071/IT9870949>
- Koste W, Shiel RJ, 1990a. Rotifera from Australian inland waters. V. Lecanidae (Rotifera: Monogononta). Transactions of the Royal Society of South Australia, 114:1-36.
- Koste W, Shiel RJ, 1990b. Rotifera from Australian inland waters. VI. Proalidae, Lindiidae (Rotifera: Monogononta). Transactions of the Royal Society of South Australia, 114:129-143.
- Koste W, Voigt M, 1978. Rotatoria. Die Rädertier Mitteleuropas. Ein Bestimmungswerk, Überordnung Monogononta. I. Textband. Gebrüder Borntraeger, Berlin, pp. 1-673.
- Kutikova LA, 1970. Kolovratki fauny USSR (Rotatoria). Akademia Nauka, Leningrad, pp. 1-742.
- National Institute of Biological Resources, 2021. National species list of Korea, 2021 [Internet]. National Institute of Biological Resources, Incheon, Accessed 10 Jan 2022, <<https://kbr.go.kr/>>.
- Republic of Korea, 2014. The fifth national report [Internet]. Convention on Biology Diversity, Montreal, Accessed 10 Jan 2022, <<https://www.cbd.int/reports/nr5/>>.
- Sanoamuang L, 1993. Comparative studies on scanning electron microscopy of trophi of the genus *Filinia* Bory De St. Vincent (Rotifera). Hydrobiologia, 264:115-128. <https://doi.org/10.1007/BF00014098>
- Sanoamuang L, 2002. Genus *Filinia* Bory de St. Vincent, 1824. In: Guides to the identification of the microinvertebrates of the continental waters of the world. 18. Rotifera. Vol. 6: Asplanchnidae, Gastropodidae, Lindiidae, Microcodidae, Synchaetidae, Trochosphaeridae (Eds., Nogrady T, Segers H). Backhuys Publishers, Leiden, pp. 224-257.
- Sanoamuang L, Segers H, Dumont HJ, 1995. Additions to the rotifer fauna of south-east Asia: new and rare species from north-east Thailand. Hydrobiologia, 313/314:35-45. <https://doi.org/10.1007/BF00025929>
- Segers H, 1995. Guides to the identification of the microinvertebrates of the continental waters of the world. 6. Rotifera Vol. 2: The Lecanidae (Monogononta). SPB Academic Publishing bv, The Hague, pp. 1-226.
- Segers H, De Smet WH, Fischer C, Fontaneto D, Michaloudi E, Wallace RL, Jersabek CD, 2012. Towards a list of Available Names in Zoology, partim Phylum Rotifera. Zootaxa, 3179:61-68. <https://doi.org/10.11646/zootaxa.3179.1.3>
- Song MO, 2015. New records of one monogonont and 5 bdelloid rotifers from Korea. Korean Journal of Environmental Biology, 33:140-147. <https://doi.org/10.11626/KJEB.2015.33.2.140>
- Song MO, 2017. New records of 13 rotifers including *Bryceella perpusilla* Wilts et al., 2010 and *Philodina lepta* Wulfert, 1951 from Korea. Journal of Species Research, 6 (Special Edition):26-37.
- Song MO, Kim HS, 1989. Monogonont Rotifers (Monogononta: Rotifera) inhabiting several lowland swamps in Kyongsangnam-do, Korea. Korean Journal of Systematic Zoology, 5:139-157.
- Tamura K, Stecher G, Kumar S, 2021. MEGA11: Molecular Evolutionary Genetics Analysis version 11. Molecular Biology and Evolution, 38:3022-3027. <https://doi.org/10.1093/molbev/msab120>
- Thrope VG, 1891. New and foreign Rotifera. Journal of the Royal Microscopical Society, 11:301-306.
- Turner PN, 1986. Some rotifers from Republic of Korea. Hydrobiologia, 137:3-7. <https://doi.org/10.1007/BF00004166>
- Wang JJ, 1961. Fauna of freshwater rotifers of China. Science Press of China, Beijing, pp. 1-288.
- Weber EF, 1898. Faune Rotatorienne du bassin du Léman. Revue Suisse de Zoologie, 5:263-785.

- Wulfert K, 1937. Zur Kenntnis der Lebensgemeinschaften der Restlochgewässer des Braunkohlenbergbaues. 1. Die Rädertiere. *Zeitschrift für Naturwissenschaften*, 91:56-69.
- Yamamoto K, 1953. Preliminary studies on the Rotatorian Fauna of Korea. *Pacific Science*, 7:151-164.
- Zhang Y, Xu S, Sun C, Dumont H, Han BP, 2021. A new set of highly efficient primers for COI amplification in rotifers.

Mitochondrial DNA Part B, 6:636-640. <https://doi.org/10.1080/23802359.2021.1878951>

Received January 7, 2022
Revised March 28, 2022
Accepted March 29, 2022